

1 **Novel skeletal effects of Glucagon-like peptide-1 (GLP-1) receptor**
2 **agonists**

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17 **Short title:** GLP-1 receptor agonists and bone

18 **Abstract**

19 Type 2 Diabetes Mellitus (T2DM) leads to bone fragility and predisposes to increased risk of
20 fracture, poor bone healing and other skeletal complications. In addition, some anti-diabetic
21 therapies for T2DM can have notable detrimental skeletal effects. Thus an appropriate
22 therapeutic strategy for T2DM should not only be effective in re-establishing good glycaemic
23 control but also in minimising skeletal complications. There is increasing evidence that
24 Glucagon-like peptide-1 receptor agonists (GLP-1RAs), now greatly prescribed for the
25 treatment of T2DM, have beneficial skeletal effects although the underlying mechanisms
26 are not completely understood. This review provides an overview of the direct and indirect
27 effects of GLP-1RAs on bone physiology, focusing on bone quality and novel mechanisms of
28 action on the vasculature and hormonal regulation. The overall experimental studies
29 indicate significant positive skeletal effects of GLP-1RAs on bone quality and strength
30 although their mechanisms of actions may differ according to various GLP-1RAs and clinical
31 studies supporting their bone protective effects are still lacking. The possibility that GLP-
32 1RAs could improve blood supply to bone, which is essential for skeletal health, is of major
33 interest and suggests that GLP-1 anti-diabetic therapy could benefit the rising number of
34 elderly T2DM patients with osteoporosis and high fracture risk.

35

36 **Lay Abstract**

37 Bone weakening is an important complication in individuals with type 2 diabetes (T2DM).
38 This review summarises the effects on skeletal health of drugs that are similar to the
39 hormone Glucagon-like peptide-1 (GLP-1), which are now used increasingly for the
40 treatment of T2DM and could lead to a stronger skeleton.

41

42 **1 Introduction**

43 Diabetes mellitus (DM) is a chronic disease that progresses worldwide at alarming rates. For
44 instance, in 2013, it has been estimated that DM affected 382 million individuals
45 (Federation, 2013). Projections for 2035 indicate a global burden of 55% to reach up to 592
46 million individuals (Federation, 2013). Associated complications are commonly cardio-
47 vascular events, nephropathy, retinopathy, neuropathy and bone fragility that dampen the
48 quality of life of affected individuals.

49 Type 2 diabetes mellitus (T2DM) is by far the most common form of DM and is characterised
50 by chronic hyperglycaemia and hyperinsulinaemia mostly caused by insulin resistance (IR) in
51 peripheral tissues such as the liver and muscle. The aetiology of bone fragility in T2DM is
52 unclear. Indeed, bone mineral density is normal or slightly elevated in T2DM despite an
53 increase risk of femoral neck fracture, suggesting alterations of bone "quality" rather than
54 bone mass (Vestergaard et al., 2005, Schwartz et al., 2011, Napoli et al., 2016). Bone quality
55 is an umbrella term that regroups factors such as bone microarchitectures, tissue material
56 properties and bone toughness (Chappard et al., 2011). Another important contributor for
57 bone fracture is represented by an increased risk in falls in this population (Schwartz et al.,
58 2002, Schwartz et al., 2008). At the cellular and molecular levels, T2DM is characterised by a
59 reduction in bone turnover suggesting modifications of bone cell behaviours (Vestergaard,
60 2007). Furthermore, low testosterone and vitamin D levels, and high plasma sclerostin, are
61 common features observed in T2DM patients (Sellmeyer et al., 2016).

62 Current treatment options of T2DM rely on lifestyle intervention and oral or injectable
63 drugs, when needed, to reach an HbA_{1c} level of 7% or less. Among the most prescribed
64 drugs, the glucagon-like peptide-1 receptor agonists (GLP-1RAs) have recently attracted
65 attention as *Glp-1r* knockout animals and GLP-1 supplemented animals exhibited
66 modifications of bone strength and quality as described below.

67 Endogenously, GLP-1 is produced by post-translational processing of the glucagon gene in
68 enteroendocrine cells, mainly L-cells (Habib et al., 2012). Two forms of GLP-1 are produced
69 in the intestine, GLP-1_{7-36NH₂} and GLP-1₇₋₃₇ although the major circulating form is GLP-1_{7-36NH₂}
70 (Orskov et al., 1994). L-cells are an open type endocrine cells highly polarised with secretory
71 granules at their basolateral pole ready to be released in the capillary network running

72 through the *lamina propria*. This secretion is regulated by intraluminal contents, neural
 73 stimuli and hormones (Baggio and Drucker, 2007). Beyond its endocrine mode of action,
 74 GLP-1 has also been suspected to act via the autonomous nervous system and hypothalamic
 75 and brainstem nuclei (Holst and Deacon, 2005).

76 To act, GLP-1 engages its receptor; the GLP-1r that is coded by the human *GLP1R* gene
 77 comprising 13 exons that span approximately 13.8kb (Yamada et al., 1995) and localised on
 78 chromosome 6p21 (Gremlich et al., 1995). The GLP-1r is expressed in the endocrine
 79 pancreas, gastro-intestinal tract, lung, heart, kidney and several regions of the brain (Baggio
 80 and Drucker, 2007). Recent evidences also suggest that GLP-1 can bind in specific
 81 circumstances to the glucagon receptor (Weston et al., 2015). The principal physiological
 82 role of GLP-1 is to potentiate glucose-dependent insulin secretion (McIntosh et al., 2010).
 83 Extrapancreatic actions of GLP-1 results in reduction of food intake through the CNS,
 84 inhibition of gastric emptying, positive actions on the cardiovascular system and a role in
 85 energy expenditure (McIntosh et al., 2010).

86 GLP-1RAs are GLP-1 with extended half-life to be more resistant to degradation by the
 87 dipeptidyl peptidase-4 (DPP-4) enzyme. Several molecules listed in Table 1 have been
 88 developed by the pharmaceutical industry and now been approved for the treatment of
 89 T2DM. The aim of the present review is to provide the reader with a comprehensive analysis
 90 of the effects of GLP-1RAs on bone physiology with special focuses on the mode of action
 91 including effects on bone quality, blood flow to bone, and on the hormonal regulation of
 92 bone metabolism.

93

94 **2 Pathogenesis of bone fragility in diabetes**

95 As mentioned in the introduction section, the aetiology of diabetes seems linked to bone
 96 quality rather than bone quantity. As such it is important to understand what alterations of
 97 bone tissue are observed in T2DM individuals.

98

2.1 Alterations in bone microarchitecture and bone material properties

Often, the assessment of bone microarchitecture and material properties require the use of bone biopsy as a source of bone tissue for experimental investigation. However, such biopsies are not available and microarchitecture and material properties have been investigated in humans by high resolution peripheral quantitative computed tomography (HR-pQCT) and bone microindentation. In term of bone microarchitecture, most studies tend to indicate a preserved trabecular bone microarchitecture but an increase in cortical bone porosity in diabetic individuals with or without fracture (Burghardt et al., 2010, Farr et al., 2014, Patsch et al., 2013). A limitation of HR-pQCT is that it can only be performed at peripheral skeletal sites and may not reflect the full bone phenotype.

In terms of bone material properties, the use of the OsteoProbe bone microindentation device showed that postmenopausal women with T2DM had significantly lower bone material strength index (BMSi) as compared to age- and sex-matched postmenopausal women without diabetes, suggesting altered bone material properties (Farr et al., 2014).

2.2 Advanced glycation endproducts

Prolonged hyperglycaemia leads to the formation of advanced glycation endproducts (AGEs) in the bone matrix that can impair its mechanical properties and the behaviour of bone cells. The most studied AGEs in humans is pentosidine because of its easiness to be measured in clinical samples such as blood or urine. As such, serum and urine pentosidine levels have been correlated with clinical fractures in T2DM patients (Schwartz et al., 2009, Yamamoto et al., 2008). However, further work is required to determine the extent to which circulating levels of AGEs reflect those in human bone tissue.

2.3 Bone turnover markers and circulating sclerostin levels

Multiple studies in humans have found that serum markers of bone formation and resorption are reduced in diabetic individuals vs. non-diabetic controls (Dobnig et al., 2006, Gerdhem et al., 2005, Krakauer et al., 1995, Shu et al., 2012). In contrast, circulating levels of sclerostin have reported to be higher in diabetic individuals (Garcia-Martin et al., 2012,

Gaudio et al., 2012, Gennari et al., 2012) and with regards to sclerostin's potent inhibitory action on bone formation, this may exacerbate the low bone formation phenotype in those patients. As such, long-standing low bone turnover observed in diabetes may result in a defective microdamage repair and increased bone microcrack accumulation that can further contribute to the observed fracture risk. Enzymatic cross-linking of type I collagen by lysyl oxidase is reduced in diabetes (Khosravi et al., 2014, Saito et al., 2006). As circulating markers of bone resorption are based on cross-linked fragments of type I collagen, it is possible that bone resorption is underestimated in diabetes.

3 Skeletal effects of GLP-1RAs: direct and/or indirect mechanisms of action

3.1 Clinical studies

Clinical data on the skeletal effects of GLP-1RAs are scarce. Bone turnover markers and bone mineral density have been assessed in T2DM patients treated with exenatide and liraglutide. However, all these studies reported no effects of GLP-1RA treatment on circulating bone markers or bone mineral density (Li et al., 2015, Bunck et al., 2011, Gilbert et al., 2016). Interestingly, the effects of liraglutide administration on bone turnover markers have been reported not in diabetic but in the obese population for the weight-loss action of liraglutide. In that study, bone formation was improved as indicated by higher values for N-terminal propeptide of type 1 procollagen reported in the liraglutide arm, but no effects on bone resorption were observed (Iepsen et al., 2015).

Two meta-analyses have also been performed on the use of GLP-1RAs and the possible effects of these medications on fracture risk. They showed divergent effects on bone fractures and differences among GLP-1RAs. It was demonstrated that liraglutide significantly reduced the risk of bone fractures whereas exenatide treatment was associated with an elevated risk of incident bone fractures (Su et al., 2015). The other meta-analysis however found neutral effect of both liraglutide and exenatide as compared with other anti-diabetic medications (Mabilleau et al., 2014). Interestingly, Driessen and colleagues (Driessen et al., 2015b, Driessen et al., 2015a) investigated in the British and Danish populations the incidence of bone fracture in GLP-1RA takers as compared with non-takers. No significant

157 difference was observed and they suggested that the effect of both GLP-1RA-type was
158 neutral in the human diabetic population.

159 However, interpretation of the above clinical studies and meta-analyses/observational
160 studies should be done carefully as they have some limitations:

- 161 • Bone fractures were not the principal end-points and as such are often disclosed as a
162 serious adverse event, although this represents only a fraction of all fractures
- 163 • There is a lack of information on bone status (bone mineral density,
164 microarchitecture, bone quality) and calcium and phosphorus metabolism at
165 baseline and at the end of studies that could highlight the possible action of GLP-
166 1RAs on bone strength
- 167 • The duration of studies may not be long enough to allow for improvement in bone
168 quality independently of bone turnover markers
- 169 • The incidence of GLP-1RAs on falls, and hence a possible mechanism of action to
170 reduce fracture, is very scarce.

171 Furthermore, as discussed below, a reduction in bone fracture has been evidenced with
172 DPP-4 inhibitors (Monami et al., 2011). However, several differences exist between DPP-4
173 inhibitors and GLP-1RAs. First, GLP-1RAs induce a modest weight loss whilst DPP-4 inhibitors
174 are neutral on that aspect (Amori et al., 2007, Inzucchi et al., 2012). After the age of 50,
175 weight loss is associated with an increased risk of fracture in overweight and obese
176 individuals (Jensen et al., 1994, Langlois et al., 2001). Secondly, the most common
177 treatment-emergent adverse events with GLP-1RAs are nausea, vomiting and diarrhoea and
178 it is plausible that they result in malabsorption of mineral and nutrients, negatively affecting
179 bone physiology.

180 In clinical trials, GLP-1RAs have been effective in reducing HbA1c level and hence chronic
181 hyperglycaemia. Data on AGEs and pentosidine in response to GLP-1RAs on the other hand
182 are limited. Tanaka et al, (Tanaka et al., 2015a) demonstrated that despite evident action of
183 liraglutide in reducing circulating glucose in Japanese overweight/obese patients with
184 T2DM, the effects of such molecule on circulating pentosidine were null.

185

186 **3.2 Effect of DPP-4 inhibitors on the skeleton**

187 The other class of pharmacotherapeutic agents that uses the incretin system are DPP-4
 188 inhibitors which inhibit the principal enzyme responsible for the degradation of endogenous
 189 GLP-1. By decreasing clearance of GLP-1, concentrations of active GLP-1 are increased by 2-
 190 to 3-fold, resulting in a lowering of fasting and postprandial glucose concentrations.

191 Data regarding the effects of DPP-4 inhibitors on human skeletal health are quite scarce. A
 192 meta-analysis carried out on 28 trials suggests a reduced fracture risk with DPP-4 inhibitors,
 193 dependent on the treatment duration (Monami et al., 2011). However, not all studies are
 194 showing a positive effect of DPP-4 inhibitors on fracture risk, BMD and bone turnover
 195 (Monami et al., 2011, Driessen et al., 2014). Recent preclinical studies showed protective
 196 effect of DPP-4 inhibitors on the skeleton of diabetic rats (Glorie et al., 2014, Eom et al.,
 197 2016) while others have shown no effect (Gallagher et al., 2014). *In vitro* studies have also
 198 indicated neutral effects of DPP-4 inhibitors on bone formation (Gallagher et al., 2014).
 199 Therefore, DPP4 inhibitors could have a possible protective effect mediated by an increase
 200 of the circulating concentrations of GLP-1 or no adverse effect. Overall, although the
 201 interest in this new anti-diabetic treatment effect on bone is high, unfortunately to date
 202 data on DPP-4 inhibitors do not allow the stating of recommendations.

203

204 **3.3 Experimental studies**

205 The first understanding of GLP-1 actions in skeletal physiology arises from *Glp1-r* KO mouse.
 206 At 10 weeks of age, these mice exhibited a small reduction in bone mass associated with an
 207 increased number of osteoclasts and eroded surfaces (Yamada et al., 2008). On the other
 208 hand, the mineral apposition and bone formation rates appeared unaffected by GLP-1r
 209 inactivation (Yamada et al., 2008). Similarly, observations in the same KO model at 16 weeks
 210 of age and in the double incretin receptor knockout model at 26 weeks of age corroborated
 211 these findings (Mabilleau, 2017, Mieczkowska et al., 2015). Taken together these results
 212 suggested a control of bone resorption (osteoclast differentiation and/or action) by the GLP-
 213 1r. According to the literature, this effect on resorption seems to be indirect through a

reduction in calcitonin gene expression in GLP-1r-deficient animals (Yamada et al., 2008) but further evidences are warranted.

While it is well established that GLP-1RAs increase bone mass in rodents (see paragraph 4), previous investigations of their effects on bone turnover are conflicting. It has been reported that 3 µg/kg/day and 4.2 µg/kg/day exenatide induced bone formation by osteoblast activation in old ovariectomised (OVX) rats (Ma et al., 2013) and in hindlimb-unloading rats (Meng et al., 2016) by promoting the osteogenic differentiation and inhibiting BMSC adipogenic differentiation. A decrease of osteoclastic surfaces was also observed (Ma et al., 2013). In contrast, we found no effect of both 10 µg/kg/day exenatide and 0.3 mg/kg/day liraglutide on bone formation and mineralisation rates in OVX mice and a slight increase of osteoclastic surfaces with the drug using bone histomorphometry (Pereira et al., 2015). The reasons for those discrepancies are unclear and may involve differences in bone turnover in mice and rats and/or in the duration of GLP-1RA treatment. Interestingly, our recent unpublished data demonstrate that GLP-1RAs increase bone formation in a T2DM mouse model but not in lean control mice, suggesting that glucose levels and/or low bone turnover may also influence the skeletal effects of GLP-1RAs. It is possible that the efficacy of GLP-1RAs on the skeleton may be improved in situations where there is a disproportionate reduction in bone formation as compared with resorption such as in T2DM.

3.4 *In vitro* studies

While several studies have reported that GLP-1RAs could have beneficial effects on the skeleton, the downstream molecular mechanisms underlying the osteogenic effect have not been identified (Bjarnason et al., 2002, Clowes et al., 2002). It is indeed unclear whether the mechanism of action of GLP-1RAs in bone is direct, through a functional GLP-1r expressed by bone cells, or indirect, via an increase in calcitonin production by the thyroid C-cell which inhibits bone resorption (Yamada et al., 2008). Furthermore, the presence and the identity of the GLP-1r in bone were controversial until recently and thus the basis for direct skeletal effects of GLP-1 has not been established.

We recently demonstrated that GLP-1 might directly affect bone cells via a GLP-1r identified in primary mouse osteoblasts isolated from calvaria and bone marrow-derived osteoclasts (Pereira et al., 2015) and this was confirmed *in situ* using a GLP-1r antibody (abcam). Similarly, other studies showed that mouse osteoblast-like MC3T3-E1 cells express a functional receptor for GLP-1 (Aoyama et al., 2014). In contrast, expression of the pancreatic-type GLP-1r mRNA was identified in human osteoblastic cell lines deriving from osteosarcomas, but its expression was dependent on the stage of osteoblastic development (Pacheco-Pantoja et al., 2011). However, other study failed to demonstrate the presence of GLP-1r at the mRNA level in primary murine osteoblasts or osteoclasts (Mabilleau et al., 2013). Similarly, the presence of the pancreatic GLP-1r in osteocytic cells was controversial as it has been reported in some cell lines, but not all (Pereira et al., 2015, Kim et al., 2013), as well as in osteocytes in rat femurs (Kim et al., 2013).

The presence of GLP-1r in bone cells *in vitro* and *in situ* implies that GLP-1RAs could have direct effects on bone cells. A study has indeed identified potential skeletal beneficial effects of 10 nM of exenatide by promoting osteoblastogenesis and restraining adipogenesis through a β -catenin pathway, during BMMSC differentiation (Meng et al., 2016). Despite increased osteoblastogenesis, no direct effect of GLP-1RAs on bone nodule mineralisation *in vitro* was shown with up to 100 nM of exenatide and 1000nM of liraglutide (Ma et al., 2013, Pereira et al., 2015). It is well established that exposure of primary osteoblast cells to high glucose levels inhibits *in vitro* bone nodule formation (Pereira et al., 2016, Balint et al., 2001). Interestingly, despite no effect of exenatide on bone formation in normal glucose conditions, unpublished results from our group demonstrate that it can reduce the deleterious effect of glucose on bone formation *in vitro*, in a dose-dependent manner. This could be due to upregulated GLP-1r expression in high glucose conditions, which could in turn magnify the effect of GLP-1RAs (Aoyama et al., 2014). Regarding the effects of GLP-1RAs on osteoclastogenesis *in vitro*, we showed that both liraglutide and exenatide increased osteoclastogenesis, while decreasing the area resorbed per osteoclast, suggesting that GLP-1RAs stimulate osteoclastic differentiation but impair their resorptive activity (Pereira et al., 2015).

3.5 GLP-1RA effects on the balance between adipogenesis and osteogenesis and adipocytes

Bone marrow mesenchymal stem cells (BMMSCs) have the ability to differentiate into various cell types, including osteoblasts and adipocytes and can be targeted by anti-diabetic drugs (e.g. thiazolidinedione). Considering the reciprocal relationship between osteogenic and adipogenic differentiation, GLP1-RAs may also indirectly affect bone formation by modulating adipogenesis. Several previous *in vitro* studies have indeed shown that GLP-1 stimulates adipose-derived stem cells (Lee et al., 2015, Cantini et al., 2015) and BMMSC (Lu et al., 2015) towards osteoblast differentiation whereas it inhibits adipocytic differentiation. Furthermore, the GLP-1R is expressed by adipocytes and GLP-1RAs down-regulates adipogenic/lipogenic genes on adipose tissue explants and cultured adipocytes while increasing lipolytic markers and expression of adiponectin (Cantini et al., 2015, El Bekay et al., 2016, Wang et al., 2017). While skeletal effects of adiponectin are multi-faceted and not always concordant, it was suggested that it may be a negative regulator of bone metabolism (Naot et al., 2017), adding to the complexity of the indirect effects of GLP-1RAs on the skeleton. Adipocyte accumulation in the bone marrow during ageing and obesity was recently shown to inhibit bone healing in mice and this was reversed by DPP-4 inhibitors, suggesting that targeting adipocytes with GLP-1RAs may also have beneficial effects on skeletal health (Ambrosi et al., 2017).

3.6 Potential signalling mechanisms of GLP-1 in bone

GLP-1RA binding to the classical pancreatic GLP-1r activates the main (cAMP-PKA) and alternative phosphoinositide 3-kinase (PI3K) and mitogen-activated protein kinase (MAPK) downstream signalling pathways. While it is still unclear whether GLP-1RAs' skeletal effects are direct via a GLP-1r expressed in bone or indirect, Cantini et al, (71) suggested that in tissues other than pancreas, GLP-1 and GLP-1RAs may not exert their actions through the classical GLP-1r but via unknown alternative pathways. This was observed in cardiomyocytes, liver and muscles (Cantini et al., 2016).

Similarly, GLP-1 action in bone could be mediated by a receptor different from the classical pancreatic cAMP-linked GLP-1r. In fact, Nuche-Berenguer et al, (72) identified in a mouse

osteoblastic line a receptor different from the pancreatic cAMP-linked GLP-1r. Moreover, they showed that GLP-1 binding to this different receptor induced an immediate hydrolysis of glycosylphosphatidylinositols, generating inositolphosphate glycan and activating PI3K and MAPK, without affecting cAMP/PKA classical signalling (Nuche-Berenguer et al., 2010b). Thus, the hypothesis of an indirect action of GLP-1RAs cannot be excluded.

4 GLP-1RAs and bone quality

Unfortunately, as neither peripheral quantitative computed tomography (pQCT) nor iliac crest bone biopsy are part of the usual care in diabetic clinical trials, human data on the effects of GLP-1RAs on all aspects of bone quality are presently missing. As such, the following summary of action of GLP-1RA is based on pre-clinical data obtained in animal models. Several animal models of either osteoporosis or T2DM have been used to assess the effects of two GLP-1RAs, exenatide and liraglutide, on bone quality and strength. However, data concerning potential bone effects of other GLP-1RAs, and especially those administered once weekly, are currently missing. Mice presenting a deletion of GLP-1r have also been generated and represented a suitable model to investigate the role of the GLP-1/GLP-r pathway in bone.

4.1 Effects of GLP-1RAs on bone strength

Our knowledge of the effects of the GLP-1/GLP-1r pathway on bone strength has been markedly improved by the use of *Glp-1r* KO mice. Indeed, although these animals are not diabetic, they exhibited a significant reduction in bone strength represented by lower ultimate load and stiffness (Mabilleau et al., 2013). Bone strength in response to the GLP-1RA exenatide has also been investigated in osteoporotic animal models generated either by ovariectomy or disuse. In ovariectomy-induced osteoporosis, the use of exenatide at a concentration as low as 1 µg/kg/day for 16 weeks, led to improvement in maximum load and stiffness as well as Young's modulus and ultimate stress, suggesting amelioration in bone microarchitecture and/or tissue material properties (Ma et al., 2013). In the rat tail suspension model, the administration of exenatide (4.2 µg/kg/day) for 4 weeks resulted in

higher value for maximum loading, stiffness, stress and Young's modulus, suggesting here again ameliorations in bone microarchitecture and/or tissue material properties (Meng et al., 2016). However, bone strength has not been measured after treatment with liraglutide.

4.2 Effects of GLP-1RAs on bone microarchitecture

In *Glp-1r* KO animals, unpublished observations from our group, revealed that these animals presented with a reduction in cancellous bone volume associated with a lower trabeculae numbers and higher trabecular spacing. These data have been confirmed by the elegant study of Yamada et al, (Yamada et al., 2008) who reported a significant reduction in cancellous bone mineral density in the same transgenic animal model. Alterations of cortical bone in this mouse model were also evidenced with lower outer bone diameter and cortical thickness (Mabilleau et al., 2013). Exenatide and liraglutide have been used as a treatment option in pre-clinical animal models of osteoporosis. They demonstrated positive effects on trabecular bone microarchitecture in the axial and appendicular skeleton evidenced by amelioration of structural parameters in lumbar vertebra and long bones as early as 4 weeks treatment. Indeed, both liraglutide and exenatide treatments resulted in higher bone volume/total volume (BV/TV) values (24% to 148%, depending on dose and treatment duration) and higher values for trabecular number (Tb.N), thickness (Tb.Th) and reduction in separation (Tb.Sp) (Ma et al., 2013, Meng et al., 2016, Pereira et al., 2015, Sun et al., 2016, Lu et al., 2015). When comparing the effects of both GLP-1RAs, liraglutide (0.3mg/kg/day) was more potent than exenatide (10µg/kg/day) (Pereira et al., 2015). The effects of GLP-1RAs on cortical microarchitecture were only observed after a minimum of 8 weeks treatment with exenatide or liraglutide, but highlighted significant increases in cortical thickness with 20µg/kg/day of exenatide or 0.6mg/kg/day of liraglutide (Lu et al., 2015, Sun et al., 2016).

In opposition to what is commonly observed in humans, animal models of T2DM exhibit significant alteration of trabecular and cortical microarchitectures. The effects of GLP-1RAs in diabetic animal models have also been reported. The use of exenatide at a regimen of 10µg/kg/day for 3 days in T2DM animals resulted in improvement in trabecular microarchitecture at the femur and lumbar spine (Nuche-Berenguer et al., 2011, Nuche-

Berenguer et al., 2010a). The use of liraglutide was also investigated in the Goto-Kakizaki T2DM rat model at a dose of 0.4mg/kg/day for 4 weeks. This regimen led to significant improvement in trabecular and cortical bone microarchitectures in the femur and lumbar vertebra (Sun et al., 2015).

The effects of liraglutide on bone microarchitecture have also been investigated in a T1DM mouse model. In this study, the administration of 0.093mg/kg/day liraglutide for 3 weeks did not demonstrate ameliorations of neither trabecular nor cortical microarchitectures (Mansur et al., 2015).

4.3 Effects of GLP-1RAs on tissue material properties

With respect to the improvement in bone strength and intrinsic properties (Young's modulus, stress), that are independent of the bone architecture, one could suspect action of GLP-1RA on tissue material properties. However, very little information has been reported. Tissue material properties represent a set of parameters that describe the modification of biochemical composition or organisation of the bone matrix at the molecular and nanoscale levels (Chappard et al., 2011). This encompasses for a thorough assessment of the mineral and collagen compartment. Most of our knowledge on the action of GLP-1 on tissue material properties is based on Glp1r KO mice. Indeed, in these animals, a significant reduction in enzymatic collagen cross-linking has been evidenced and associated with alteration of bone strength at the tissue level (Mabilleau et al., 2013). However, in opposition to what has been seen with the sister incretin hormone GIP, Glp-1r deletion did not alter the mineral compartment (Mieczkowska et al., 2013). Data regarding the potential effects of GLP-1RAs on tissue material properties in osteoporotic animals are lacking. However, an elegant study conducted by Mansur et al, (Mansur et al., 2015) investigated the effects of 0.093 mg/kg/day liraglutide over a period of 3 weeks in a T1DM mouse model. These authors reported no amelioration of enzymatic collagen cross-linking or collagen glycation but an unexpected reduction in collagen destruction (Mansur et al., 2015).

5 GLP-1RAs and blood flow to bone

Diabetes leads to poor circulation and vascular diseases are the principal causes of death and disability in people with diabetes. Consequently, wound and fracture healing are delayed in diabetic patients, one of the main reasons being the impairment in vascularisation (Falanga, 2005, Loder, 1988). Particularly, diabetes was shown to induce a decrease in endothelial progenitor cells (EPC) that are important for angiogenesis and vascular repair (Fadini et al., 2005, Rigato et al., 2015). It is now well established that blood flow is crucial to bone vascular function and osteogenesis (Ramasamy et al., 2016) and that disrupted blood supply to bone is associated with reduced bone mass, osteonecrosis and impaired bone regeneration (Loder, 1988, Vogt et al., 1997, Atsumi and Kuroki, 1992). Very little work has however examined whether the bone vasculature and bone blood flow are reduced in diabetic bone and if it is possible to restore them with the use of anti-diabetic drugs. Fajardo (Fajardo, 2017) recently reviewed the literature regarding the microvascular complications in diabetic bone but evidences are still lacking to support the link between skeletal fragility in diabetes and those vascular complications.

Incretin-based therapy seems very promising for the prevention of diabetic vascular complications (Mima, 2016). The potential for GLP-1RAs to enhance vascular function has been demonstrated in several studies (Nystrom et al., 2004, Zhou et al., 2015, Sufiun et al., 2015, Smits et al., 2015). The improvement of vascular endothelial function restores impaired glucose tolerance by ameliorating insulin resistance in skeletal muscle (Kubota et al., 2011). Interestingly, two weeks administration of 0.5µg/kg/d exenatide was shown to accelerate diabetic wound healing by increasing angiogenesis in the wound and the number of circulating EPCs (Roan et al., 2017). Our recent, not yet published, work also demonstrates that 10µg/kg exenatide can have beneficial effects on bone vascularisation in diabetic bone by acutely increasing blood flow to bone in db/db mice. This suggests that the increased bone formation induced by exenatide treatment in diabetic mice could be attributed in part to this increased skeletal perfusion. No study has yet examined the effect of liraglutide on bone blood flow. More work is therefore needed to examine whether skeletal perfusion is linked to bone formation in diabetic bone and if GLP-1RAs could be used as treatment to increase vascularisation in diabetic patients with poor fracture healing.

420

421 **6 GLP-1RAs and hormones that regulate bone metabolism**

422 A major breakthrough in the bone research field has been the finding that bone is an
 423 endocrine organ that can affect other organs via the release of hormones such as
 424 osteocalcin and sclerostin. There are increasing reports showing that GLP-1RAs can affect
 425 the release of these hormones by bone cells *in vitro* and in animal models but the clinical
 426 evidence is however still very scarce.

427

428 **6.1 Sclerostin**

429 The discovery of the importance of the Wnt/ β catenin pathway for bone formation has led
 430 to extensive work examining the function of sclerostin in bone. Sclerostin is a product of the
 431 *SOST* gene expressed mainly by osteocytes which is secreted and acts as a potent antagonist
 432 of Wnt signalling (Bellido, 2014). Its deficiency or its pharmacological neutralisation
 433 increases bone formation, making it a potential target for treatment of bone diseases
 434 associated with bone loss, such as osteoporosis (Hamann et al., 2013, Ominsky et al., 2010).
 435 Most studies have shown that serum sclerostin levels are elevated in diabetic patients,
 436 suggesting that sclerostin could contribute to the decreased bone formation observed in
 437 diabetic patients (Garcia-Martin et al., 2012, Gaudio et al., 2012, Gennari et al., 2012).
 438 However the association between sclerostin levels and increased fracture risk in T2DM
 439 patients is not always conclusive and further studies are required to confirm the link (Yu et
 440 al., 2017). Some differences in serum sclerostin levels measurements could be explained by
 441 the fact that sclerostin could be derived from other non-skeletal sources so that serum
 442 levels may not always reflect the production in bone (Roforth et al., 2014) and also because
 443 the ELISA kits for sclerostin measurements were found to lack accuracy (Piec et al., 2016,
 444 Costa et al., 2017).

445 To address this issue, experimental studies were conducted examining if sclerostin
 446 production by osteocytes is modified in bone of diabetic rodents or *in vitro* when osteocytes
 447 are cultured in high glucose levels. Although an *in vitro* study reported an increased
 448 production of sclerostin by osteocyte-like cell line when cultured in hyperglycemia (Pereira

et al., 2016, Tanaka et al., 2015b), our recent study shows that the impaired bone microarchitecture and cellular turnover associated with T2DM-like conditions in diabetic ZDF rats are not correlated with changes in serum sclerostin levels, bone sclerostin expression or osteocyte viability (Pereira et al., 2016). On the other hand, high fat diet in mice resulted in increased serum sclerostin and dramatic alterations of osteocyte network organisation (Mabilleau et al., 2016).

Few studies have investigated if GLP-1RAs could affect sclerostin production by osteocytes. Although GLP-1r is mainly expressed by immature osteoblasts, it can be present in osteocytes where it co-localise with sclerostin (Kim et al., 2013), suggesting that GLP-1RAs may affect sclerostin production. Kim et al, (Kim et al., 2013) have indeed shown that sclerostin levels are increased in diabetic rats compared to controls and can be down-regulated by exenatide treatment. More recently, they demonstrate that the DPP-4 inhibitor vildagliptin lowers the increased levels of sclerostin induced by thiazolidinedione (Eom et al., 2016). Our own results showed that exenatide but not liraglutide decreased sclerostin levels in OVX mice (Pereira et al., 2015).

Overall, despite some controversy, the majority of clinical and experimental studies suggest that sclerostin may play a role in the decreased bone turnover in patients with T2DM and be a potential target for GLP-1 therapy. The origin of sclerostin in serum is however still unclear and more studies are required to examine whether bone production of sclerostin is affected in T2DM patients.

6.2 Osteocalcin

Osteocalcin (OC) is a small protein produced in bone by osteoblasts during bone formation which has traditionally been used as a serum marker for bone formation (Ducy et al., 1996). This protein has however regained a different interest in recent years due to the demonstration that when it is in its uncarboxylated form (GluOC) which does not bind to bone, it can circulate, act as a hormone and regulate glucose metabolism (Lee et al., 2007). GluOC can stimulate the release of GLP-1 from the small intestine and therefore indirectly promote insulin secretion by the pancreatic β -cell (Mizokami et al., 2013).

It was suggested that incretins could contribute to whole body energy metabolism by modulating osteocalcin synthesis in osteoblasts. The effects of GLP-1RAs on osteocalcin production by osteoblasts were examined and once again the results are inconsistent. While Kim et al, (Kim et al., 2013), Nuche-Berenguer et al, (Nuche-Berenguer et al., 2010a) demonstrate an increase in serum osteocalcin levels with exenatide in T2DM and IR rats, this was not the case with liraglutide treatment (Iepsen et al., 2015, Conte et al., 2015). A recent study demonstrates that incretins inhibit thyroid hormone-stimulated osteocalcin synthesis in osteoblasts *in vitro*, suggesting that incretins could stimulate bone formation by reducing the osteocalcin levels (Kainuma et al., 2016), although this is not confirmed *in vivo*. Osteocalcin concentration significantly increases during calcification and arterial calcification is an important complication of diabetes due to the differentiation of vascular smooth muscle cells into osteoblast-like cells. Although some work demonstrates an inhibitory effect of GLP-1RAs on vascular calcifications, this is not always the case (Zhan et al., 2014, Davenport et al., 2015).

6.3 Calcitonin

Calcitonin is a peptide hormone produced by the thyroid parafollicular cells, commonly named "C-cells," that regulate calcium homeostasis (Warshawsky et al., 1980). Increases in serum calcium activate the release of calcitonin from the C-cells, which consecutively inhibits bone resorption by the osteoclast and calcium absorption by the intestine. It was therefore one of the first agents to be used as a treatment for osteoporosis. As mentioned previously, several animal studies suggest that GLP-1RAs can affect bone metabolism indirectly via the release of calcitonin by thyroid C cells which express the GLP-1r (Yamada et al., 2008, Lamari et al., 1996). The expression of the GLP-1r in thyroid glands has indeed been documented in rodents (Bjerre Knudsen et al., 2010), but there is an uncertainty regarding its expression in humans (Gier et al., 2012, Hegedus et al., 2011). Furthermore, basal and stimulated calcitonin did not change during 1 year of liraglutide treatment (Lunati et al., 2016). Our own work demonstrates that serum levels of calcitonin were indeed increased by exenatide treatment in ovariectomised mice (Pereira et al., 2015), although this was not shown when mice were treated with liraglutide, suggesting once again that these two GLP-1 agonists may have a different mechanism of action.

7 Differences in mechanisms of action between liraglutide and exenatide

Some divergent skeletal effects of liraglutide and exenatide observed in clinical and experimental studies suggest possible different mechanisms of action. Although overall similar in action, liraglutide and exenatide treatments differ in several aspects mainly due to their differences in molecular structures. Indeed, liraglutide is an analog of human naïve GLP-1 with 97 % homology, whereas exenatide only share 50% homology with human naïve GLP-1. This molecular divergence determines the differences in pharmacokinetic profiles between liraglutide and exenatide (Jespersen et al., 2013). The half-life of liraglutide is five time longer than exenatide, therefore exenatide treatment requires twice daily injections in patients. Moreover, while exenatide is mainly eliminated in the kidney, liraglutide is fully degraded within the body and no specific organ or enzyme is responsible for its elimination (Giorda et al., 2014). Exenatide administration also results in higher frequency of antibody formation than that of liraglutide (Buse et al., 2011). Thus, the favourable role of liraglutide on bone fractures risk and its more potent effect *in vivo* could be explained in part by its similar pharmacokinetic profile with human naïve GLP-1. On the other hand, exenatide possesses distinct absorption, elimination and antibody formation properties. Whether those different properties of exenatide could interact with some bone metabolism and turnover pathways needs to be clearly elucidated but this may explain the distinctive effects of these two GLP-1RAs on bone hormones production.

8 Conclusion

Based on several rodent studies, GLP-1 therapy emerges as one of the most promising anti-diabetic therapy for treating the skeletal fragility associated with diabetes. It was shown to increase bone mass, improve trabecular and cortical architectures, enhance bone strength and tissue material properties, affecting the collagen compartment rather than the mineral one. The possible mechanisms of action of GLP-1RAs on the skeleton are illustrated in Figure 1. They are however still not very clear and different GLP-1RAs may have different means of action. Among the potential ones, the stimulation of bone blood flow by GLP-1RAs seems very interesting and extremely promising in situations of osteoporotic and diabetic fractures. Clinical data are however still lacking and those establishing the relationship

between the GLP-1RA use and decrease fracture risk have been so far negative. There is therefore a need for long-term clinical studies comparing the skeletal effects of different GLP-1RAs.

Declaration of interest

There is no conflict of interest that could be perceived as prejudicing the impartiality of the research presented

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References

AMBROSI, T. H., SCIALDONE, A., GRAJA, A., GOHLKE, S., JANK, A. M., BOCIAN, C., WOELK, L., FAN, H., LOGAN, D. W., SCHURMANN, A., et al. 2017. Adipocyte Accumulation in the Bone Marrow during Obesity and Aging Impairs Stem Cell-Based Hematopoietic and Bone Regeneration. *Cell Stem Cell*, 20, 771-784 e6.

AMORI, R. E., LAU, J. & PITTAS, A. G. 2007. Efficacy and safety of incretin therapy in type 2 diabetes: systematic review and meta-analysis. *JAMA*, 298, 194-206.

AOYAMA, E., WATARI, I., PODYMA-INOUE, K. A., YANAGISHITA, M. & ONO, T. 2014. Expression of glucagon-like peptide-1 receptor and glucosedependent insulintropic polypeptide receptor is regulated by the glucose concentration in mouse osteoblastic MC3T3-E1 cells. *Int J Mol Med*, 34, 475-82.

ATSUMI, T. & KUROKI, Y. 1992. Role of impairment of blood supply of the femoral head in the pathogenesis of idiopathic osteonecrosis. *Clin Orthop Relat Res*, 22-30.

BAGGIO, L. L. & DRUCKER, D. J. 2007. Biology of incretins: GLP-1 and GIP. *Gastroenterology*, 132, 2131-57.

- 565 BALINT, E., SZABO, P., MARSHALL, C. F. & SPRAGUE, S. M. 2001. Glucose-induced inhibition
566 of in vitro bone mineralization. *Bone*, 28, 21-8.
- 567 BELLIDO, T. 2014. Osteocyte-driven bone remodeling. *Calcif Tissue Int*, 94, 25-34.
- 568 BJARNASON, N. H., HENRIKSEN, E. E., ALEXANDERSEN, P., CHRISTGAU, S., HENRIKSEN, D. B.
569 & CHRISTIANSEN, C. 2002. Mechanism of circadian variation in bone resorption. *Bone*, 30,
570 307-13.
- 571 BJERRE KNUDSEN, L., MADSEN, L. W., ANDERSEN, S., ALMHOLT, K., DE BOER, A. S.,
572 DRUCKER, D. J., GOTFREDSEN, C., EGEROD, F. L., HEGELUND, A. C., JACOBSEN, H., et al.
573 2010. Glucagon-like Peptide-1 receptor agonists activate rodent thyroid C-cells causing
574 calcitonin release and C-cell proliferation. *Endocrinology*, 151, 1473-86.
- 575 BUNCK, M. C., ELIASSON, B., CORNER, A., HEINE, R. J., SHAGINIAN, R. M., TASKINEN, M. R.,
576 YKI-JARVINEN, H., SMITH, U. & DIAMANT, M. 2011. Exenatide treatment did not affect bone
577 mineral density despite body weight reduction in patients with type 2 diabetes. *Diabetes*
578 *Obes Metab*, 13, 374-7.
- 579 BURGHARDT, A. J., ISSEVER, A. S., SCHWARTZ, A. V., DAVIS, K. A., MASHARANI, U.,
580 MAJUMDAR, S. & LINK, T. M. 2010. High-resolution peripheral quantitative computed
581 tomographic imaging of cortical and trabecular bone microarchitecture in patients with type
582 2 diabetes mellitus. *J Clin Endocrinol Metab*, 95, 5045-55.
- 583 BUSE, J. B., GARBER, A., ROSENSTOCK, J., SCHMIDT, W. E., BRETT, J. H., VIDEBAEK, N.,
584 HOLST, J. & NAUCK, M. 2011. Liraglutide treatment is associated with a low frequency and
585 magnitude of antibody formation with no apparent impact on glycemic response or
586 increased frequency of adverse events: results from the Liraglutide Effect and Action in
587 Diabetes (LEAD) trials. *J Clin Endocrinol Metab*, 96, 1695-702.
- 588 CANTINI, G., DI FRANCO, A., SAMAVAT, J., FORTI, G., MANNUCCI, E. & LUCONI, M. 2015.
589 Effect of liraglutide on proliferation and differentiation of human adipose stem cells. *Mol*
590 *Cell Endocrinol*, 402, 43-50.
- 591 CANTINI, G., MANNUCCI, E. & LUCONI, M. 2016. Perspectives in GLP-1 Research: New
592 Targets, New Receptors. *Trends Endocrinol Metab*, 27, 427-38.

- 593 CHAPPARD, D., BASLE, M. F., LEGRAND, E. & AUDRAN, M. 2011. New laboratory tools in the
594 assessment of bone quality. *Osteoporos Int*, 22, 2225-40.
- 595 CLOWES, J. A., HANNON, R. A., YAP, T. S., HOYLE, N. R., BLUMSOHN, A. & EASTELL, R. 2002.
596 Effect of feeding on bone turnover markers and its impact on biological variability of
597 measurements. *Bone*, 30, 886-90.
- 598 CONTE, C., CECERE, A., GUGLIELMI, G. & NAPOLI, N. 2015. Letter to the Editor: "GLP-1
599 Receptor Agonist Treatment Increases Bone Formation and Prevents Bone Loss in Weight-
600 Reduced Obese Women" by Iepsen E.W., et al. *J Clin Endocrinol Metab*, 100, L92-3.
- 601 COSTA, A. G., CREMERS, S. & BILEZIKIAN, J. P. 2017. Sclerostin measurement in human
602 disease: Validity and current limitations. *Bone*, 96, 24-28.
- 603 DAVENPORT, C., MAHMOOD, W. A., FORDE, H., ASHLEY, D. T., AGHA, A., MCDERMOTT, J.,
604 SREENAN, S., THOMPSON, C. J., MCGRATH, F., MCADAM, B., et al. 2015. The effects of
605 insulin and liraglutide on osteoprotegerin and vascular calcification in vitro and in patients
606 with type 2 diabetes. *Eur J Endocrinol*, 173, 53-61.
- 607 DOBNIG, H., PISWANGER-SOLKNER, J. C., ROTH, M., OBERMAYER-PIETSCH, B., TIRAN, A.,
608 STRELE, A., MAIER, E., MARITSCHNEGG, P., SIEBERER, C. & FAHRLEITNER-PAMMER, A. 2006.
609 Type 2 diabetes mellitus in nursing home patients: effects on bone turnover, bone mass,
610 and fracture risk. *J Clin Endocrinol Metab*, 91, 3355-63.
- 611 DRIESSEN, J. H., HENRY, R. M., VAN ONZENOORT, H. A., LALMOHAMED, A., BURDEN, A. M.,
612 PRIETO-ALHAMBRA, D., NEEF, C., LEUFKENS, H. G. & DE VRIES, F. 2015a. Bone fracture risk is
613 not associated with the use of glucagon-like peptide-1 receptor agonists: a population-
614 based cohort analysis. *Calcif Tissue Int*, 97, 104-12.
- 615 DRIESSEN, J. H., VAN ONZENOORT, H. A., HENRY, R. M., LALMOHAMED, A., VAN DEN BERGH,
616 J. P., NEEF, C., LEUFKENS, H. G. & DE VRIES, F. 2014. Use of dipeptidyl peptidase-4 inhibitors
617 for type 2 diabetes mellitus and risk of fracture. *Bone*, 68, 124-30.
- 618 DRIESSEN, J. H., VAN ONZENOORT, H. A., STARUP-LINDE, J., HENRY, R., BURDEN, A. M.,
619 NEEF, C., VAN DEN BERGH, J. P., VESTERGAARD, P. & DE VRIES, F. 2015b. Use of Glucagon-

- 620 Like-Peptide 1 Receptor Agonists and Risk of Fracture as Compared to Use of Other Anti-
621 hyperglycemic Drugs. *Calcif Tissue Int*, 97, 506-15.
- 622 DUCY, P., DESBOIS, C., BOYCE, B., PINERO, G., STORY, B., DUNSTAN, C., SMITH, E., BONADIO,
623 J., GOLDSTEIN, S., GUNDBERG, C., et al 1996. Increased bone formation in osteocalcin-
624 deficient mice. *Nature*, 382, 448-52.
- 625 EL BEKAY, R., COIN-ARAGUEZ, L., FERNANDEZ-GARCIA, D., OLIVA-OLIVERA, W., BERNAL-
626 LOPEZ, R., CLEMENTE-POSTIGO, M., DELGADO-LISTA, J., DIAZ-RUIZ, A., GUZMAN-RUIZ, R.,
627 VAZQUEZ-MARTINEZ, R., et al. 2016. Effects of glucagon-like peptide-1 on the differentiation
628 and metabolism of human adipocytes. *Br J Pharmacol*, 173, 1820-34.
- 629 EOM, Y. S., GWON, A. R., KWAK, K. M., KIM, J. Y., YU, S. H., LEE, S., KIM, Y. S., PARK, I. B.,
630 KIM, K. W., LEE, K., et al. 2016. Protective Effects of Vildagliptin against Pioglitazone-Induced
631 Bone Loss in Type 2 Diabetic Rats. *PLoS One*, 11, e0168569.
- 632 FADINI, G. P., MIORIN, M., FACCO, M., BONAMICO, S., BAESSO, I., GREGO, F., MENEGOLO,
633 M., DE KREUTZENBERG, S. V., TIENGO, A., AGOSTINI, C., et al. Circulating endothelial
634 progenitor cells are reduced in peripheral vascular complications of type 2 diabetes mellitus.
635 *J Am Coll Cardiol*, 45, 1449-57.
- 636 FAJARDO, R. J. 2017. Is Diabetic Skeletal Fragility Associated with Microvascular
637 Complications in Bone? *Curr Osteoporos Rep*, 15, 1-8.
- 638 FALANGA, V. 2005. Wound healing and its impairment in the diabetic foot. *Lancet*, 366,
639 1736-43.
- 640 FARR, J. N., DRAKE, M. T., AMIN, S., MELTON, L. J., 3RD, MCCREADY, L. K. & KHOSLA, S. 2014.
641 In vivo assessment of bone quality in postmenopausal women with type 2 diabetes. *J Bone*
642 *Miner Res*, 29, 787-95.
- 643 FEDERATION, I. D. 2013. *IDF Diabetes Atlas*.
- 644 GALLAGHER, E. J., SUN, H., KORNHAUSER, C., TOBIN-HESS, A., EPSTEIN, S., YAKAR, S. &
645 LEROITH, D. 2014. The effect of dipeptidyl peptidase-IV inhibition on bone in a mouse model
646 of type 2 diabetes. *Diabetes Metab Res Rev*, 30, 191-200.

- 647 GARCIA-MARTIN, A., ROZAS-MORENO, P., REYES-GARCIA, R., MORALES-SANTANA, S.,
 648 GARCIA-FONTANA, B., GARCIA-SALCEDO, J. A. & MUNOZ-TORRES, M. 2012. Circulating levels
 649 of sclerostin are increased in patients with type 2 diabetes mellitus. *J Clin Endocrinol Metab*,
 650 97, 234-41.
- 651 GAUDIO, A., PRIVITERA, F., BATTAGLIA, K., TORRISI, V., SIDOTI, M. H., PULVIRENTI, I.,
 652 CANZONIERI, E., TRINGALI, G. & FIORE, C. E. 2012. Sclerostin levels associated with inhibition
 653 of the Wnt/beta-catenin signaling and reduced bone turnover in type 2 diabetes mellitus. *J*
 654 *Clin Endocrinol Metab*, 97, 3744-50.
- 655 GENNARI, L., MERLOTTI, D., VALENTI, R., CECCARELLI, E., RUVIO, M., PIETRINI, M. G.,
 656 CAPODARCA, C., FRANCI, M. B., CAMPAGNA, M. S., CALABRO, A., et al. 2012. Circulating
 657 sclerostin levels and bone turnover in type 1 and type 2 diabetes. *J Clin Endocrinol Metab*,
 658 97, 1737-44.
- 659 GERDHEM, P., ISAKSSON, A., AKESSON, K. & OBRANT, K. J. 2005. Increased bone density and
 660 decreased bone turnover, but no evident alteration of fracture susceptibility in elderly
 661 women with diabetes mellitus. *Osteoporos Int*, 16, 1506-12.
- 662 GIER, B., BUTLER, P. C., LAI, C. K., KIRAKOSSIAN, D., DENICOLA, M. M. & YEH, M. W. 2012.
 663 Glucagon like peptide-1 receptor expression in the human thyroid gland. *J Clin Endocrinol*
 664 *Metab*, 97, 121-31.
- 665 GILBERT, M. P., MARRE, M., HOLST, J. J., GARBER, A., BAERES, F. M., THOMSEN, H. &
 666 PRATLEY, R. E. 2016. Comparison of the Long-Term Effects of Liraglutide and Glimepiride
 667 Monotherapy on Bone Mineral Density in Patients with Type 2 Diabetes. *Endocr Pract*, 22,
 668 406-11.
- 669 GIORDA, C. B., NADA, E. & TARTAGLINO, B. 2014. Pharmacokinetics, safety, and efficacy of
 670 DPP-4 inhibitors and GLP-1 receptor agonists in patients with type 2 diabetes mellitus and
 671 renal or hepatic impairment. A systematic review of the literature. *Endocrine*, 46, 406-19.
- 672 GLORIE, L., BEHETS, G. J., BAERTS, L., DE MEESTER, I., D'HAESE, P. C. & VERHULST, A. 2014.
 673 DPP IV inhibitor treatment attenuates bone loss and improves mechanical bone strength in
 674 male diabetic rats. *Am J Physiol Endocrinol Metab*, 307, E447-55.

675 GREMLICH, S., PORRET, A., HANI, E. H., CHERIF, D., VIONNET, N., FROGUEL, P. & THORENS,
 676 B. 1995. Cloning, functional expression, and chromosomal localization of the human
 677 pancreatic islet glucose-dependent insulintropic polypeptide receptor. *Diabetes*, 44, 1202-
 678 8.

679 HABIB, A. M., RICHARDS, P., CAIRNS, L. S., ROGERS, G. J., BANNON, C. A., PARKER, H. E.,
 680 MORLEY, T. C., YEO, G. S., REIMANN, F. & GRIBBLE, F. M. 2012. Overlap of endocrine
 681 hormone expression in the mouse intestine revealed by transcriptional profiling and flow
 682 cytometry. *Endocrinology*, 153, 3054-65.

683 HAMANN, C., RAUNER, M., HOHNA, Y., BERNHARDT, R., METTELSIEFEN, J., GOETTSCH, C.,
 684 GUNTHER, K. P., STOLINA, M., HAN, C. Y., ASUNCION, F. J., et al. C. 2013. Sclerostin antibody
 685 treatment improves bone mass, bone strength, and bone defect regeneration in rats with
 686 type 2 diabetes mellitus. *J Bone Miner Res*, 28, 627-38.

687 HEGEDUS, L., MOSES, A. C., ZDRAVKOVIC, M., LE THI, T. & DANIELS, G. H. 2011. GLP-1 and
 688 calcitonin concentration in humans: lack of evidence of calcitonin release from sequential
 689 screening in over 5000 subjects with type 2 diabetes or nondiabetic obese subjects treated
 690 with the human GLP-1 analog, liraglutide. *J Clin Endocrinol Metab*, 96, 853-60.

691 HOLST, J. J. & DEACON, C. F. 2005. Glucagon-like peptide-1 mediates the therapeutic actions
 692 of DPP-IV inhibitors. *Diabetologia*, 48, 612-5.

693 IEPSSEN, E. W., LUNDGREN, J. R., HARTMANN, B., PEDERSEN, O., HANSEN, T., JORGENSEN, N.
 694 R., JENSEN, J. E., HOLST, J. J., MADSBAD, S. & TOREKOV, S. S. 2015. GLP-1 Receptor Agonist
 695 Treatment Increases Bone Formation and Prevents Bone Loss in Weight-Reduced Obese
 696 Women. *J Clin Endocrinol Metab*, 100, 2909-17.

697 INZUCCHI, S. E., BERGENSTAL, R. M., BUSE, J. B., DIAMANT, M., FERRANNINI, E., NAUCK, M.,
 698 PETERS, A. L., TSAPAS, A., WENDER, R., MATTHEWS, D. R., AMERICAN DIABETES, A. &
 699 EUROPEAN ASSOCIATION FOR THE STUDY OF, D. 2012. Management of hyperglycemia in
 700 type 2 diabetes: a patient-centered approach: position statement of the American Diabetes
 701 Association (ADA) and the European Association for the Study of Diabetes (EASD). *Diabetes*
 702 *Care*, 35, 1364-79.

703 JENSEN, L. B., QUADE, F. & SORESEN, O. H. 1994. Bone loss accompanying voluntary
704 weight loss in obese humans. *J Bone Miner Res*, 9, 459-63.

705 JESPERSEN, M. J., KNOP, F. K. & CHRISTENSEN, M. 2013. GLP-1 agonists for type 2 diabetes:
706 pharmacokinetic and toxicological considerations. *Expert Opin Drug Metab Toxicol*, 9, 17-29.

707 KAINUMA, S., TOKUDA, H., FUJITA, K., KAWABATA, T., SAKAI, G., MATSUSHIMA-NISHIWAKI,
708 R., HARADA, A., KOZAWA, O. & OTSUKA, T. 2016. Attenuation by incretins of thyroid
709 hormone-stimulated osteocalcin synthesis in osteoblasts. *Biomed Rep*, 5, 771-775.

710 KHOSRAVI, R., SODEK, K. L., FAIBISH, M. & TRACKMAN, P. C. 2014. Collagen advanced
711 glycation inhibits its Discoidin Domain Receptor 2 (DDR2)-mediated induction of lysyl
712 oxidase in osteoblasts. *Bone*, 58, 33-41.

713 KIM, J. Y., LEE, S. K., JO, K. J., SONG, D. Y., LIM, D. M., PARK, K. Y., BONEWALD, L. F. & KIM, B.
714 J. 2013. Exendin-4 increases bone mineral density in type 2 diabetic OLETF rats potentially
715 through the down-regulation of SOST/sclerostin in osteocytes. *Life Sci*, 92, 533-40.

716 KRAKAUER, J. C., MCKENNA, M. J., BUDERER, N. F., RAO, D. S., WHITEHOUSE, F. W. &
717 PARFITT, A. M. 1995. Bone loss and bone turnover in diabetes. *Diabetes*, 44, 775-82.

718 KUBOTA, T., KUBOTA, N., KUMAGAI, H., YAMAGUCHI, S., KOZONO, H., TAKAHASHI, T.,
719 INOUE, M., ITOH, S., TAKAMOTO, I., SASAKO, T., et al. 2011. Impaired insulin signaling in
720 endothelial cells reduces insulin-induced glucose uptake by skeletal muscle. *Cell Metab*, 13,
721 294-307.

722 LAMARI, Y., BOISSARD, C., MOUKHTAR, M. S., JULLIENNE, A., ROSSELIN, G. & GAREL, J. M.
723 1996. Expression of glucagon-like peptide 1 receptor in a murine C cell line: regulation of
724 calcitonin gene by glucagon-like peptide 1. *FEBS Lett*, 393, 248-52.

725 LANGLOIS, J. A., MUSSOLINO, M. E., VISSER, M., LOOKER, A. C., HARRIS, T. & MADANS, J.
726 2001. Weight loss from maximum body weight among middle-aged and older white women
727 and the risk of hip fracture: the NHANES I epidemiologic follow-up study. *Osteoporos Int*, 12,
728 763-8.

- 729 LEE, H. M., JOO, B. S., LEE, C. H., KIM, H. Y., OCK, J. H. & LEE, Y. S. 2015. Effect of Glucagon-
 730 like Peptide-1 on the Differentiation of Adipose-derived Stem Cells into Osteoblasts and
 731 Adipocytes. *J Menopausal Med*, 21, 93-103.
- 732 LEE, N. K., SOWA, H., HINOI, E., FERRON, M., AHN, J. D., CONFAVREUX, C., DACQUIN, R.,
 733 MEE, P. J., MCKEE, M. D., JUNG, D. Y., et al. 2007. Endocrine regulation of energy
 734 metabolism by the skeleton. *Cell*, 130, 456-69.
- 735 LI, R., XU, W., LUO, S., XU, H., TONG, G., ZENG, L., ZHU, D. & WENG, J. 2015. Effect of
 736 exenatide, insulin and pioglitazone on bone metabolism in patients with newly diagnosed
 737 type 2 diabetes. *Acta Diabetol*, 52, 1083-91.
- 738 LODER, R. T. 1988. The influence of diabetes mellitus on the healing of closed fractures. *Clin*
 739 *Orthop Relat Res*, 210-6.
- 740 LU, N., SUN, H., YU, J., WANG, X., LIU, D., ZHAO, L., SUN, L., ZHAO, H., TAO, B. & LIU, J. 2015.
 741 Glucagon-like peptide-1 receptor agonist Liraglutide has anabolic bone effects in
 742 ovariectomized rats without diabetes. *PLoS One*, 10, e0132744.
- 743 LUNATI, M. E., GRANCINI, V., COLOMBO, C., PALMIERI, E., RESI, V., PERRINO, M., ORSI, E. &
 744 FUGAZZOLA, L. 2016. Basal and stimulated calcitonin levels in patients with type 2 diabetes
 745 did not change during 1 year of Liraglutide treatment. *Metabolism*, 65, 1-6.
- 746 MA, X., MENG, J., JIA, M., BI, L., ZHOU, Y., WANG, Y., HU, J., HE, G. & LUO, X. 2013. Exendin-
 747 4, a glucagon-like peptide-1 receptor agonist, prevents osteopenia by promoting bone
 748 formation and suppressing bone resorption in aged ovariectomized rats. *J Bone Miner Res*,
 749 28, 1641-52.
- 750 MABILLEAU, G. 2017. Interplay between bone and incretin hormones: A review.
 751 *Morphologie*, 101, 9-18.
- 752 MABILLEAU, G., MIECZKOWSKA, A. & CHAPPARD, D. 2014. Use of glucagon-like peptide-1
 753 receptor agonists and bone fractures: a meta-analysis of randomized clinical trials. *J*
 754 *Diabetes*, 6, 260-6.

755 MABILLEAU, G., MIECZKOWSKA, A., IRWIN, N., FLATT, P. R. & CHAPPARD, D. 2013. Optimal
 756 bone mechanical and material properties require a functional glucagon-like peptide-1
 757 receptor. *J Endocrinol*, 219, 59-68.

758 MABILLEAU, G., PERROT, R., FLATT, P. R., IRWIN, N. & CHAPPARD, D. 2016. High fat-fed
 759 diabetic mice present with profound alterations of the osteocyte network. *Bone*, 90, 99-106.

760 MANSUR, S. A., MIECZKOWSKA, A., BOUVARD, B., FLATT, P. R., CHAPPARD, D., IRWIN, N. &
 761 MABILLEAU, G. 2015. Stable Incretin Mimetics Counter Rapid Deterioration of Bone Quality
 762 in Type 1 Diabetes Mellitus. *J Cell Physiol*, 230, 3009-18.

763 MCINTOSH, C. H., WIDENMAIER, S. & KIM, S. J. 2010. Pleiotropic actions of the incretin
 764 hormones. *Vitam Horm*, 84, 21-79.

765 MENG, J., MA, X., WANG, N., JIA, M., BI, L., WANG, Y., LI, M., ZHANG, H., XUE, X., HOU, Z., et
 766 al. 2016. Activation of GLP-1 Receptor Promotes Bone Marrow Stromal Cell Osteogenic
 767 Differentiation through beta-Catenin. *Stem Cell Reports*, 6, 633.

768 MIECZKOWSKA, A., IRWIN, N., FLATT, P. R., CHAPPARD, D. & MABILLEAU, G. 2013. Glucose-
 769 dependent insulintropic polypeptide (GIP) receptor deletion leads to reduced bone
 770 strength and quality. *Bone*, 56, 337-42.

771 MIECZKOWSKA, A., MANSUR, S., BOUVARD, B., FLATT, P. R., THORENS, B., IRWIN, N.,
 772 CHAPPARD, D. & MABILLEAU, G. 2015. Double incretin receptor knock-out (DIRKO) mice
 773 present with alterations of trabecular and cortical micromorphology and bone strength.
 774 *Osteoporos Int*, 26, 209-18.

775 MIMA, A. 2016. Incretin-Based Therapy for Prevention of Diabetic Vascular Complications. *J*
 776 *Diabetes Res*, 2016, 1379274.

777 MIZOKAMI, A., YASUTAKE, Y., GAO, J., MATSUDA, M., TAKAHASHI, I., TAKEUCHI, H. &
 778 HIRATA, M. 2013. Osteocalcin induces release of glucagon-like peptide-1 and thereby
 779 stimulates insulin secretion in mice. *PLoS One*, 8, e57375.

780 MONAMI, M., DICEMBRINI, I., ANTENORE, A. & MANNUCCI, E. 2011. Dipeptidyl peptidase-4
 781 inhibitors and bone fractures: a meta-analysis of randomized clinical trials. *Diabetes Care*,
 782 34, 2474-6.

- 783 NAOT, D., MUSSON, D. S. & CORNISH, J. 2017. The Activity of Adiponectin in Bone. *Calcif*
784 *Tissue Int*, 100, 486-499.
- 785 NAPOLI, N., CHANDRAN, M., PIERROZ, D. D., ABRAHAMSEN, B., SCHWARTZ, A. V., FERRARI,
786 S. L., BONE, I. O. F. & DIABETES WORKING, G. 2016. Mechanisms of diabetes mellitus-
787 induced bone fragility. *Nat Rev Endocrinol*.
- 788 NUCHE-BERENGUER, B., LOZANO, D., GUTIERREZ-ROJAS, I., MORENO, P., MARINOSO, M. L.,
789 ESBRIT, P. & VILLANUEVA-PENACARRILLO, M. L. 2011. GLP-1 and exendin-4 can reverse
790 hyperlipidic-related osteopenia. *J Endocrinol*, 209, 203-10.
- 791 NUCHE-BERENGUER, B., MORENO, P., PORTAL-NUNEZ, S., DAPIA, S., ESBRIT, P. &
792 VILLANUEVA-PENACARRILLO, M. L. 2010a. Exendin-4 exerts osteogenic actions in insulin-
793 resistant and type 2 diabetic states. *Regul Pept*, 159, 61-6.
- 794 NUCHE-BERENGUER, B., PORTAL-NUNEZ, S., MORENO, P., GONZALEZ, N., ACITORES, A.,
795 LOPEZ-HERRADON, A., ESBRIT, P., VALVERDE, I. & VILLANUEVA-PENACARRILLO, M. L. 2010b.
796 Presence of a functional receptor for GLP-1 in osteoblastic cells, independent of the cAMP-
797 linked GLP-1 receptor. *J Cell Physiol*, 225, 585-92.
- 798 NYSTROM, T., GUTNIAK, M. K., ZHANG, Q., ZHANG, F., HOLST, J. J., AHREN, B. & SJOHOLM,
799 A. 2004. Effects of glucagon-like peptide-1 on endothelial function in type 2 diabetes
800 patients with stable coronary artery disease. *Am J Physiol Endocrinol Metab*, 287, E1209-15.
- 801 OMINSKY, M. S., VLASSEROS, F., JOLETTE, J., SMITH, S. Y., STOUCH, B., DOELLGAST, G.,
802 GONG, J., GAO, Y., CAO, J., GRAHAM, K., et al. 2010. Two doses of sclerostin antibody in
803 cynomolgus monkeys increases bone formation, bone mineral density, and bone strength. *J*
804 *Bone Miner Res*, 25, 948-59.
- 805 ORSKOV, C., RABENHOJ, L., WETTERGREN, A., KOFOD, H. & HOLST, J. J. 1994. Tissue and
806 plasma concentrations of amidated and glycine-extended glucagon-like peptide I in humans.
807 *Diabetes*, 43, 535-9.
- 808 PACHECO-PANTOJA, E. L., RANGANATH, L. R., GALLAGHER, J. A., WILSON, P. J. & FRASER, W.
809 D. 2011. Receptors and effects of gut hormones in three osteoblastic cell lines. *BMC Physiol*,
810 11, 12.

811 PATSCH, J. M., BURGHARDT, A. J., YAP, S. P., BAUM, T., SCHWARTZ, A. V., JOSEPH, G. B. &
 812 LINK, T. M. 2013. Increased cortical porosity in type 2 diabetic postmenopausal women with
 813 fragility fractures. *J Bone Miner Res*, 28, 313-24.

814 PEREIRA, M., GOHIN, S., LUND, N., HVID, A., SMITHAM, P. J., ODDY, M. J., REICHERT, I.,
 815 FARLAY, D., ROUX, J. P., CLEASBY, M. E. et al. 2016. Sclerostin does not play a major role in
 816 the pathogenesis of skeletal complications in type 2 diabetes mellitus. *Osteoporosis*
 817 *International*, 1-12.

818 PEREIRA, M., JEYABALAN, J., JORGENSEN, C. S., HOPKINSON, M., AL-JAZZAR, A., ROUX, J. P.,
 819 CHAVASSIEUX, P., ORRISS, I. R., CLEASBY, M. E. & CHENU, C. 2015. Chronic administration of
 820 Glucagon-like peptide-1 receptor agonists improves trabecular bone mass and architecture
 821 in ovariectomised mice. *Bone*, 81, 459-67.

822 PIEC, I., WASHBOURNE, C., TANG, J., FISHER, E., GREEVES, J., JACKSON, S. & FRASER, W. D.
 823 2016. How Accurate is Your Sclerostin Measurement? Comparison Between Three
 824 Commercially Available Sclerostin ELISA Kits. *Calcif Tissue Int*, 98, 546-55.

825 RAMASAMY, S. K., KUSUMBE, A. P., SCHILLER, M., ZEUSCHNER, D., BIXEL, M. G., MILIA, C.,
 826 GAMREKELASHVILI, J., LIMBOURG, A., MEDVINSKY, A., SANTORO, M. M., et al. 2016. Blood
 827 flow controls bone vascular function and osteogenesis. *Nat Commun*, 7, 13601.

828 RIGATO, M., BITTANTE, C., ALBIERO, M., AVOGARO, A. & FADINI, G. P. 2015. Circulating
 829 Progenitor Cell Count Predicts Microvascular Outcomes in Type 2 Diabetic Patients. *J Clin*
 830 *Endocrinol Metab*, 100, 2666-72.

831 ROAN, J. N., CHENG, H. N., YOUNG, C. C., LEE, C. J., YEH, M. L., LUO, C. Y., TSAI, Y. S. & LAM,
 832 C. F. 2017. Exendin-4, a glucagon-like peptide-1 analogue, accelerates diabetic wound
 833 healing. *J Surg Res*, 208, 93-103.

834 ROFORTH, M. M., FUJITA, K., MCGREGOR, U. I., KIRMANI, S., MCCREADY, L. K., PETERSON, J.
 835 M., DRAKE, M. T., MONROE, D. G. & KHOSLA, S. 2014. Effects of age on bone mRNA levels of
 836 sclerostin and other genes relevant to bone metabolism in humans. *Bone*, 59, 1-6.

837 SAITO, M., FUJII, K., MORI, Y. & MARUMO, K. 2006. Role of collagen enzymatic and glycation
 838 induced cross-links as a determinant of bone quality in spontaneously diabetic WBN/Kob
 839 rats. *Osteoporos Int*, 17, 1514-23.

840 SCHWARTZ, A. V., GARNERO, P., HILLIER, T. A., SELLMEYER, D. E., STROTMEYER, E. S.,
 841 FEINGOLD, K. R., RESNICK, H. E., TYLAVSKY, F. A., BLACK, D. M., CUMMINGS, S. R., et al.
 842 2009. Pentosidine and increased fracture risk in older adults with type 2 diabetes. *J Clin*
 843 *Endocrinol Metab*, 94, 2380-6.

844 SCHWARTZ, A. V., HILLIER, T. A., SELLMEYER, D. E., RESNICK, H. E., GREGG, E., ENSRUD, K. E.,
 845 SCHREINER, P. J., MARGOLIS, K. L., CAULEY, J. A., NEVITT, M. C., et al. 2002. Older women
 846 with diabetes have a higher risk of falls: a prospective study. *Diabetes Care*, 25, 1749-54.

847 SCHWARTZ, A. V., VITTINGHOFF, E., BAUER, D. C., HILLIER, T. A., STROTMEYER, E. S.,
 848 ENSRUD, K. E., DONALDSON, M. G., CAULEY, J. A., HARRIS, T. B., KOSTER, A., et al. 2011.
 849 Association of BMD and FRAX score with risk of fracture in older adults with type 2 diabetes.
 850 *JAMA*, 305, 2184-92.

851 SCHWARTZ, A. V., VITTINGHOFF, E., SELLMEYER, D. E., FEINGOLD, K. R., DE REKENEIRE, N.,
 852 STROTMEYER, E. S., SHORR, R. I., VINIK, A. I., ODDEN, M. C., PARK, S. W., et al. 2008.
 853 Diabetes-related complications, glycemic control, and falls in older adults. *Diabetes Care*, 31,
 854 391-6.

855 SELLMEYER, D. E., CIVITELLI, R., HOFBAUER, L. C., KHOSLA, S., LECKA-CZERNIK, B. &
 856 SCHWARTZ, A. V. 2016. Skeletal Metabolism, Fracture Risk, and Fracture Outcomes in Type
 857 1 and Type 2 Diabetes. *Diabetes*, 65, 1757-66.

858 SHU, A., YIN, M. T., STEIN, E., CREMERS, S., DWORAKOWSKI, E., IVES, R. & RUBIN, M. R.
 859 2012. Bone structure and turnover in type 2 diabetes mellitus. *Osteoporos Int*, 23, 635-41.

860 SMITS, M. M., MUSKIET, M. H., TONNEIJCK, L., KRAMER, M. H., DIAMANT, M., VAN RAALTE,
 861 D. H. & SERNE, E. H. 2015. GLP-1 Receptor Agonist Exenatide Increases Capillary Perfusion
 862 Independent of Nitric Oxide in Healthy Overweight Men. *Arterioscler Thromb Vasc Biol*, 35,
 863 1538-43.

- 864 SU, B., SHENG, H., ZHANG, M., BU, L., YANG, P., LI, L., LI, F., SHENG, C., HAN, Y., QU, S. &
 865 WANG, J. 2015. Risk of bone fractures associated with glucagon-like peptide-1 receptor
 866 agonists' treatment: a meta-analysis of randomized controlled trials. *Endocrine*, 48, 107-15.
- 867 SUFIUN, A., RAFIQ, K., FUJISAWA, Y., RAHMAN, A., MORI, H., NAKANO, D., KOBORI, H.,
 868 OHMORI, K., MASAKI, T., KOHNO, M. et al. 2015. Effect of dipeptidyl peptidase-4 inhibition
 869 on circadian blood pressure during the development of salt-dependent hypertension in rats.
 870 *Hypertens Res*, 38, 237-43.
- 871 SUN, H. X., LU, N., LIU, D. M., ZHAO, L., SUN, L. H., ZHAO, H. Y., LIU, J. M. & TAO, B. 2016. The
 872 bone-preserving effects of exendin-4 in ovariectomized rats. *Endocrine*, 51, 323-32.
- 873 SUN, H. X., LU, N., LUO, X., ZHAO, L. & LIU, J. M. 2015. Liraglutide, the glucagon-like peptide-
 874 1 receptor agonist, has anabolic bone effects in diabetic Goto-Kakizaki rats. *J Diabetes*, 7,
 875 584-8.
- 876 TANAKA, K., SAISHO, Y., KAWAI, T., TANAKA, M., MEGURO, S., IRIE, J., IMAI, T., SHIGIHARA,
 877 T., MORIMOTO, J., YAJIMA, K., et al. 2015a. Efficacy and safety of liraglutide monotherapy
 878 compared with metformin in Japanese overweight/obese patients with type 2 diabetes.
 879 *Endocr J*, 62, 399-409.
- 880 TANAKA, K., YAMAGUCHI, T., KANAZAWA, I. & SUGIMOTO, T. 2015b. Effects of high glucose
 881 and advanced glycation end products on the expressions of sclerostin and RANKL as well as
 882 apoptosis in osteocyte-like MLO-Y4-A2 cells. *Biochem Biophys Res Commun*, 461, 193-9.
- 883 VESTERGAARD, P. 2007. Discrepancies in bone mineral density and fracture risk in patients
 884 with type 1 and type 2 diabetes--a meta-analysis. *Osteoporos Int*, 18, 427-44.
- 885 VESTERGAARD, P., REJNMARK, L. & MOSEKILDE, L. 2005. Relative fracture risk in patients
 886 with diabetes mellitus, and the impact of insulin and oral antidiabetic medication on relative
 887 fracture risk. *Diabetologia*, 48, 1292-9.
- 888 VOGT, M. T., CAULEY, J. A., KULLER, L. H. & NEVITT, M. C. 1997. Bone mineral density and
 889 blood flow to the lower extremities: the study of osteoporotic fractures. *J Bone Miner Res*,
 890 12, 283-9.

- 891 WANG, A., LI, T., AN, P., YAN, W., ZHENG, H., WANG, B. & MU, Y. 2017. Exendin-4
892 Upregulates Adiponectin Level in Adipocytes via Sirt1/Foxo-1 Signaling Pathway. *PLoS One*,
893 12, e0169469.
- 894 WARSHAWSKY, H., GOLTZMAN, D., ROULEAU, M. F. & BERGERON, J. J. 1980. Direct in vivo
895 demonstration by radioautography of specific binding sites for calcitonin in skeletal and
896 renal tissues of the rat. *J Cell Biol*, 85, 682-94.
- 897 WESTON, C., LU, J., LI, N., BARKAN, K., RICHARDS, G. O., ROBERTS, D. J., SKERRY, T. M.,
898 POYNER, D., PARDAMWAR, M., REYNOLDS, C. A., et al. 2015. Modulation of Glucagon
899 Receptor Pharmacology by Receptor Activity-modifying Protein-2 (RAMP2). *J Biol Chem*,
900 290, 23009-22.
- 901 YAMADA, C., YAMADA, Y., TSUKIYAMA, K., YAMADA, K., UDAGAWA, N., TAKAHASHI, N.,
902 TANAKA, K., DRUCKER, D. J., SEINO, Y. & INAGAKI, N. 2008. The murine glucagon-like
903 peptide-1 receptor is essential for control of bone resorption. *Endocrinology*, 149, 574-9.
- 904 YAMADA, Y., HAYAMI, T., NAKAMURA, K., KAISAKI, P. J., SOMEYA, Y., WANG, C. Z., SEINO, S.
905 & SEINO, Y. 1995. Human gastric inhibitory polypeptide receptor: cloning of the gene (GIPR)
906 and cDNA. *Genomics*, 29, 773-6.
- 907 YAMAMOTO, M., YAMAGUCHI, T., YAMAUCHI, M., YANO, S. & SUGIMOTO, T. 2008. Serum
908 pentosidine levels are positively associated with the presence of vertebral fractures in
909 postmenopausal women with type 2 diabetes. *J Clin Endocrinol Metab*, 93, 1013-9.
- 910 YU, O. H., RICHARDS, B., BERGER, C., JOSSE, R. G., LESLIE, W. D., GOLTZMAN, D., KAISER, S.
911 M., KOVACS, C. S. & DAVISON, K. S. 2017. The association between sclerostin and incident
912 type 2 diabetes risk: a cohort study. *Clin Endocrinol (Oxf)*, 86, 520-525.
- 913 ZHAN, J. K., TAN, P., WANG, Y. J., WANG, Y., HE, J. Y., TANG, Z. Y., HUANG, W. & LIU, Y. S.
914 2014. Exenatide can inhibit calcification of human VSMCs through the NF-kappaB/RANKL
915 signaling pathway. *Cardiovasc Diabetol*, 13, 153.
- 916 ZHOU, X., HUANG, C. H., LAO, J., POCAI, A., FORREST, G., PRICE, O., ROY, S., KELLEY, D. E.,
917 SULLIVAN, K. A. & FORREST, M. J. 2015. Acute hemodynamic and renal effects of glucagon-

918 like peptide 1 analog and dipeptidyl peptidase-4 inhibitor in rats. *Cardiovasc Diabetol*, 14,
919 29.
920

Figure 1: Simplified scheme of the possible skeletal effects of GLP-1RAs

GLP-1RAs exert multiple beneficial effects on the skeleton. They increase bone mass, improve trabecular and cortical architectures, enhance bone strength and collagen content. They however do not affect bone mineral density (BMD). Several potential mechanisms of action have been described to explain these positive effects of GLP-1RAs on bone. They include indirect effects of GLP-1RAs on bone turnover mediated via hormonal changes. GLP-1RAs were indeed shown to upregulate calcitonin production by C-cells in the thyroid leading to a decrease in bone resorption; alternatively they can down-regulate sclerostin production by osteocytes and increase bone formation. Their beneficial effects on bone blood flow could also contribute to a stimulation of bone formation. GLP-1RAs can also have direct effects on bone cells mediated by the GLP-1R expressed in primary osteoblasts, osteoclasts and in some osteocytes. *In vitro* studies suggest that GLP-1RAs may stimulate bone formation in condition of hyperglycemia and impair osteoclast bone resorptive activity. However, some divergent skeletal effects of liraglutide and exenatide were observed in clinical and experimental studies, suggesting that different GLP-1RAs may use various mechanisms of action.

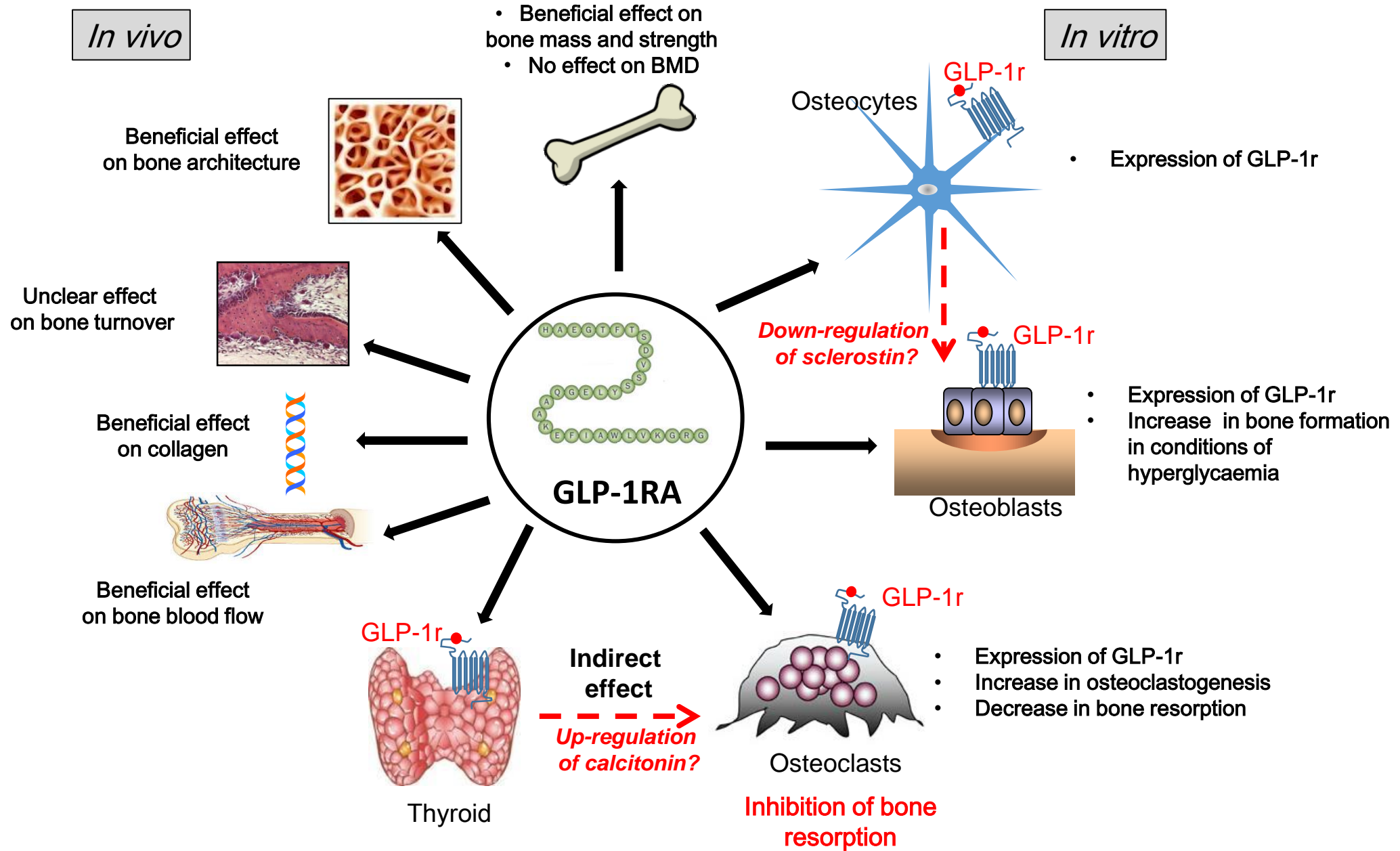


Table 1: Summary of approved GLP-1RAs for the treatment of type 2 diabetes mellitus

Active compound	Drug name	Marketed by	Approved in	Approved dose range
Exenatide (or Exendin-4)	Byetta	Astra Zeneca AB	2006	5-10 µg twice daily
Liraglutide	Victoza	Novo Nordisk A/S	2009	0.6-1.8 mg once daily
Lixisenatide	Lyxumia	Sanofi Aventis Groupe	2013	10-20 µg once daily
Exenatide (or Exendin-4) long acting release	Bydureon	Astra Zeneca AB	2011	2 mg once weekly
Albiglutide	Eperzan	GlaxoSmithKline Trading Services Ltd	2014	30-50 mg once weekly
Dulaglutide	Trulicity	Eli Lilly Nederland B.V.	2014	0.75-1.5 mg once weekly